## **PCT**

# WORLD INTELLECTUAL PROPERTY ORGANIZATION International Bureau



### INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification <sup>6</sup> : A61L 27/00	A1	(11) International Publication Number: WO 98/44965
		(43) International Publication Date: 15 October 1998 (15.10.98)
(21) International Application Number: PCT/GB (22) International Filing Date: 6 April 1998 ( (30) Priority Data: 9706967.8 5 April 1997 (05.04.97) 9712399.6 14 June 1997 (14.06.97)  (71) Applicant (for all designated States except US): C LIMITED [GB/GB]; 12 North Harbour Estate, 8AA (GB).  (72) Inventor; and (75) Inventor/Applicant (for US only): HEALY, David, [IE/GB]; Midton House, Alloway, Ayr KA7 4EG  (74) Agent: MURGITROYD & COMPANY; 373 Scotlar Glasgow G5 8QA (GB).	GILTEC Ayr KA , Micha (GB).	BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE GH, GM, GW, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasiar patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), Europear patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).  Published  With international search report.  Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

### (54) Title: IMPLANTATION COMPOSITION COMPRISING GLASS PARTICLES

#### (57) Abstract

There is provided a composition suitable for implantation in soft tissue (for example at or around a body orifice) in order to augment the volume of soft tissue. The composition described comprises particles of a, preferably water-soluble, biodegradable glass in a suitable carrier medium, such as glycerol. The particles, which are desirably irregularly shaped, may have an average particle diameter of from 50  $\mu$ m to 2000  $\mu$ m, preferably 50  $\mu$ m to 300  $\mu$ m. By injecting the particles into soft tissue, for example the bladder submucosa, it is possible to bulk up the soft tissue where this is required. This procedure can be applied to treat conditions such as vesicoureteric reflux. Additionally the procedure could be used cosmetically.

## FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
ΑT	Austria	FR	France	LU	Luxembourg	SN	Senegal
ΑU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
ΑZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav	TM	Turkmenistan
BF	Burkina Faso	GR	Greece		Republic of Macedonia	TR	Turkey
BG	Bulgaria	HU	Hungary	ML	Mali	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MN	Mongolia	UA	Ukraine
BR	Brazil	IL	Israel	MR	Mauritania	UG	Uganda
BY	Belarus	IS	Keland	MW	Malawi	US	United States of America
CA	Canada	IT	Italy	MX	Mexico	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NE	Niger	VN	Viet Nam
CG	Congo	KE	Kenya	NL	Netherlands	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NO	Norway	zw	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's	NZ	New Zealand		
CM	Cameroon		Republic of Korea	PL	Poland		
CN	China	KR	Republic of Korea	PT	Portugal		
CU	Cuba	KZ	Kazakstan	RO	Romania		
CZ	Czech Republic	LC	Saint Lucia	RU	Russian Federation		
DE	Germany	u	Liechtenstein	SD	Sudan		
DK	Denmark	LK	Sri Lanka	SE	Sweden		
EE	Estonia	LR	Liberia	SG	Singapore		

1

1 IMPLANTATION COMPOSITION COMPRISING GLASS PARTICLES 2 3 The present invention is concerned with a composition suitable for implantation at or in the vicinity of a 4 5 body orifice or sphincter muscle to aid correct 6 function. 7 8 Many body functions rely upon the correct functioning of sphincter muscles. For example, the pyloric 9 10 sphincter controls when the contents of the stomach 11 pass into the small intestine. Similarly, the urethral 12 sphincter controls when the contents of the bladder are 13 Incorrect functioning due to premature 14 relaxation of such sphincter muscles can be 15 problematic, and in the case of stress urinary 16 incontinence (malfunction of the urethral sphincter) 17 highly distressing to the patient. 18 19 Premature relaxation of a sphincter muscle often occurs 20 when the sphincter muscle itself lacks sufficient bulk 21 to adequately close the orifice in question. 22 option to overcome the problem is by implanting bulking 23 material in the submucosa surrounding the orifice, 24 thereby reducing the area to be closed by the sphincter 25 Generally, the bulking material is injected

1 into the site to augment the soft tissue present. 2 Suitable bulking materials are available commercially 3 and are generally in the form of spherical particles or 4 beads based on silicone, PTFE or collagen. These beads 5 are suspended in a carrier fluid such as glycerine or 6 The carrier fluid acts as a lubricant during hydrogel. 7 the implantation process and assists expulsion of the 8 implant from the syringe through an endoscopic needle. 9 The carrier fluid is eliminated from the body and the 10 implant material gradually becomes encapsulated by 11 collagen at the implant site. The collagen capsule 12 which forms around the implanted material adds to the 13 bulk at the site. One such bulking material is 14 MACROPLASTIQUE (Trade Mark) of Uroplasty, Inc. 15 16 Existing implants do not biodegrade but remain 17 permanently in the body of the patient. Recently, 18 concern has been raised that such implants may 19 gradually migrate away from the site of implantation 20 during the lifetime of the patient. Thus, the original 21 problem may recur as the size of the implant gradually 22 decreases due to migration of the beads inserted. 23 patient will therefore need to undergo a further 24 procedure in order to insert more beads at the site 25 concerned. The migrating implant may, in addition, 26 cause irritation and such implants have been reported 27 to be associated with cancer, auto-immune and 28 connective tissue disease. 29 30 In addition to stress urinary incontinence, such 31 implants have also been used to prevent vesicoureteral 32 Vesicoureteric reflux is a condition occurring reflux. 33 in babies and small children where the ureteral orifice 34 is incompletely closed during contraction of the 35 bladder. Urine is thus allowed to reflux back up the 36 ureter and can cause recurrent infections of the

1 kidneys, frequently leading to permanent kidney damage. 2 In a similar manner to stress urinary incontinence, it is possible to insert pellets or beads of silicone 3 4 rubber or teflon in the submucosa of the bladder wall close to the ureteral orifice. Again, the procedure 5 6 requires the permanent insertion of the implant. 7 8 Paediatric vesicoureteral reflux usually resolves 9 itself as the bladder wall thickens. By the time a 10 child is five years old the urinary system has usually 11 matured sufficiently to make the implant material 12 redundant. Again, it is possible for implant material 13 to migrate from the implant site causing obstruction, 14 occlusion or embolism at another site. Implants have 15 also been associated with cancer, auto-immune and connective tissue disease. 16 17 18 The present invention provides a composition suitable 19 for implantation in soft tissue (for example at or 20 around a body orifice) in order to augment the volume of soft tissue. The composition of the present 21 22 invention comprises particles of biodegradable glass in 23 a suitable carrier medium. The carrier medium is 24 required to ensure easy injection at the site of 25 interest. 26 27 The currently available silicone, PTFE and collagen 28 beads are all deformable. This property aids injection 29 of the beads, but also contributes to their ability to 30 migrate from the site of interest. By contrast, the 31 glass particles of the present invention are non-32 deformable. 33 34 The composition is suitable for insertion in the 35 bladder submucosa to treat stress urinary incontinence 36 or vesioureteric reflux by bulking up the area around

4

1 the urethral sphincter or urethral orifice 2 respectively. 3 4 Optionally, the glass particles dissolve over a 5 relatively long period, typically one to five years, 6 more usually one to two years. 7 8 Preferably, the glass particles are irregularly shaped. 9 This contrasts to the commercially available implants 10 which are formed from spherically shaped beads. 11 irregular shape of the glass particles encourages their 12 encapsulation in fibrous tissue. Such encapsulation 13 further reduces the rate of dissolution of the glass 14 and also helps to prevent migration of the particles. 15 16 Typically, the glass particles used in the present 17 invention may have a diameter of from 50 µm up to 2000 18 More conveniently, however, the average diameter 19 of the particles will be 1000 µm or less, usually 500 20 µm or less. Good results have been obtained with 21 particles having an average diameter of 300 to 200 µm 22 or less, for example 150 µm or less. 23 24 Particles having smaller diameters, e.g. 100 µm or 25 less, particularly of approximately 50 µm, or even less, 26 are of especial interest. 27 28 One advantage of the present invention is that it is 29 possible to form glass particles having such small 30 diameters (e.g.  $50-100 \mu m$ ). Where such small particles 31 are used the problems associated with injection are 32 reduced. Additionally, once the particles have been 33 located in the site of interest, the outside surfaces 34 of the particles becomes tacky as the particles begin 35 to dissolve into body fluids so that the particles 36 become associated in situ in a sticky cohesive mass.

5

1 Such particle association greatly reduces the rate of 2 particle migration and the health risks associated 3 No such association has been observed with 4 the prior art silicone, PTFE or collagen beads. 5 6 A carrier medium is generally used to assist injection 7 of the particles. The carrier medium is typically 8 glycerol, but other conventional carrier mediums (e.g. 9 corn oil, sesame oil, sunflower oil or olibas oil) may also be used. A surfactant and/or suspending agent may 10 11 also be included in the composition. surfactants include, for example, benzyl benzoate, 12 13 ethyl oleate and benzyl alcohol. Typical suspending 14 agents include, for example, carboxymethylcellulose and 15 alginate. 16 17 In a further aspect the present invention provides a 18 method of augmenting an area of soft tissue in a body (e.g. thickening a wall of a body organ), said method 19 20 comprising injecting a composition into the soft tissue 21 (e.g. the submucosa of said wall), said composition 22 comprising particles of a biodegradable glass. 23 24 Thus, the present invention provides a method of 25 combatting vesicoureteric reflux by injecting a 26 composition of the present invention into the bladder 27 submucosa close to the ureteral orifice such that urine 28 is substantially unable to pass up the ureter upon 29 contraction of the bladder. 30 31 Likewise, if the composition of the present invention 32 is injected into the submucosa in the vicinity of the 33 urethral sphincter, stress urinary incontinence may be 34 overcome due to the "bulking" effect of the injected

35 36 particles.

6 1 The present invention may be used at other body areas 2 where soft tissue augmentation has a beneficial effect. 3 Examples include injection around the anal passage, in 4 order to reduce blood flow at the site and hence combat 5 development of haemorrhoids (piles). Likewise soft 6 tissue augmentation may be beneficial to temporarily 7 correct an "incompetent" cervix which would prevent 8 sustainment of a pregnancy. The soft tissue 9 augmentation of the present invention may further be 10 used to build up portions of the body damaged by 11 accident or surgery, allowing healing to take place. 12 Particular mention may be made of reshaping the facial 13 area of a patient. From the above examples it is clear 14 that the composition of the present invention may be 15 used not only to treat existing conditions but also for prophylactic and cosmetic purposes. 16 17 18 Generally the glass will be a controlled release glass 19 (CRG). CRGs are vitreous inorganic polymers which 20 dissolve over a pre-programmed period leaving virtually 21 no residue. The components of manufacture are all 22 present as natural body constituents hence CRGs show 23 little or no cytotoxicity and exhibit a minimal tissue 24 reaction. 11 The use of glasses which can dissolve in water and body 12

25

13

14

15

16

fluid are well-known. These glasses are formed from phosphorus pentoxide and may be modified to dissolve over a period of months or years, as required. date, such glasses have been used, in medicine, for the controlled release of a number of agents, for example, drugs, hormones and trace elements.

17 18

19 It is known that certain glasses, in which the usual 20 glass former, silicon dioxide, of traditional glasses 21 is replaced with phosphorus pentoxide as the glass

PCT/GB98/01017

7

former, are soluble in water and body fluids. 1 The rate 2 of dissolution is controlled largely by the addition of 3 glass modifiers such as calcium and magnesium oxide. 4 In simple terms, the greater the concentration of the 5 modifier the slower the rate of dissolution. 6 of dissolution which can be imparted to the glasses may range from minutes to months or even to several years. 7 It is known to include in such compositions quantities 8 9 of trace elements such as copper, cobalt and selenium 10 which will be released from the glass as it slowly 11 dissolves over the selected period of time. 12 13 The use of water-soluble glasses has been described for 14 a variety of purposes in the literature. For example, 15 UK Patent Specifications Nos 1,565,906, 2,079,152, 2,077,585 and 2,146,531 describe the gradual 16 17 dissolution of the glasses as providing a means of 18 controlled release of drugs, hormones, fungicides, 19 insecticides, spermicides and other agents with which 20 the glasses have been impregnated. The glasses are 21 used, for example, in the form of an implant or bolus. 22 23 UK Patent Specification No 2,030,559 describes the use 24 of selenium-impregnated water-soluble glass for 25 providing controlled release of the selenium as a trace element into cattle and sheep, the glass being applied 26 27 as a subcutaneous insert. UK Patent Specification 28 No 2,037,735 also describes a subcutaneous implant of 29 water-soluble glass, and in this case the glass is 30 impregnated with copper; minor quantities of trace 31 elements such as boron, arsenic, iodine, manganese, 32 chromium, silver, gold and gallium may also be 33 included. 34 35

Water-soluble glass has also been proposed for use in

1 prosthetics, for example in UK Patent Specification 2 No 2,099,702, and for use in anticorrosive paints, as described in UK Patent Specification No 2,062,612. 3 4 Further the literature provides for the use of such glasses in the controlled release of ferrous and ferric 5 6 ions into the human or animal body by ingestion or 7 implantation of the glass (UK Patent Specification 8 No 2,081,703), and for the use of glasses in the 9 controlled release of ions such as lithium, sodium, potassium, caesium, rubidium, polyphosphate, calcium 10 11 and aluminium to patients by inclusion of the glass in 12 a drip feed line (UK Patent Specification No 2,057,420). 13 14 15 Optionally the water-soluble glass may be a silver 16 containing water-soluble glass. Advantageously the 17 silver content may be introduced into the glass 18 composition in the form of silver orthophosphate. 19 20 Suitable glasses include, for example, the ARGLAES™ 21 glass of Giltech Limited. 22 23 The glass may be adapted by the use of glass modifiers 24 to give a sustained release of silver ions over a set 25 period. 26 27 In one embodiment the water-soluble glass comprises an 28 alkali metal oxide M2O, an alkaline earth oxide MO, 29 phosphorus pentoxide  $P_2O_5$  and silver oxide  $(Ag_2O)$  or 30 silver orthophosphate (Ag<sub>3</sub>PO<sub>4</sub>). 31 32 Most preferably, said glass contains not more than 40 33 mole % M<sub>2</sub>O or MO, not less than 10 mole % M<sub>2</sub>O or MO, and 34 not more than 50 mole % nor less than 38 mole % 35 phosphorus pentoxide, optionally with the inclusion of

1 0.05 to 5.0 mole % silver oxide or orthophosphate. 2 3 Said alkali metal oxide may be sodium oxide (Na<sub>2</sub>0), 4 potassium (K<sub>2</sub>0) or a mixture thereof; and said alkaline 5 earth oxide may be calcium oxide (CaO), magnesium oxide 6 (Mg0), zinc oxide (Zn0) or a mixture thereof. 7 8 The glass may also contain less than 5 mole % silicon 9 dioxide (SiO<sub>2</sub>), boric oxide (B<sub>2</sub>O<sub>3</sub>), sulphate ion (SO<sub>4</sub><sup>2-</sup>), 10 a halide ion, copper oxide (CuO) or a mixture thereof. 11 12 Typically the soluble glasses used in this invention 13 comprise phosphorus pentoxide  $(P_2O_5)$  as the principal 14 glass-former, together with any one or more 15 glass-modifying non-toxic materials such as sodium oxide  $(Na_20)$ , potassium oxide  $(K_20)$ , magnesium oxide 16 17 (Mg0), zinc oxide (Zn0) and calcium oxide (Ca0). 18 rate at which the silver-release glass dissolves in 19 fluids is determined by the glass composition, 20 generally by the ratio of glass-modifier to 21 glass-former and by the relative proportions of the 22 glass-modifiers in the glass. By suitable adjustment 23 of the glass composition, the dissolution rates in 24 water at 38 °C ranging from substantially zero to 25 25 mg/cm<sup>2</sup>/hour or more can be designed. However, the most 26 desirable dissolution rate R of the glass is between 27 0.01 and 2.0 mg/cm<sup>2</sup>/hour. The water-soluble glass is 28 preferably a phosphate glass, and the silver may 29 advantageously be introduced during manufacture as 30 silver orthophosphate (Ag<sub>3</sub>PO<sub>4</sub>). The content of silver 31 and other constituents in the glass can vary in accordance with conditions of use and desired rates of 32 release, the content of silver generally being up to 5 33 34 mole %. While we are following convention in 35 describing the composition of the glass in terms of the

10

mole % of oxides, of halides and of sulphate ions, this 1 2 is not intended to imply that such chemical species are 3 present in the glass nor that they are used for the 4 batch for the preparation of the glass. 5 The glass may be formed by a number of methods. 6 7 simply be cast by conventional or centrifugal 8 procedures, or it may be prepared via one or more 9 stages of rod, fibre or tube drawing. 10 preparation techniques include foamed glass. 11 glass formation it will be comminuted into finely 12 divided form. 13 14 Optionally, the composition of the present invention 15 may contain an active ingredient. The term "active 16 ingredient" is used herein to refer to any agent which affects the metabolism or any metabolic or cellular 17 process of the patient (including growth factors and 18 living cells), promotes healing, combats infection, 19 20 hypergranulation or inflammation. Antibiotics and 21 other anti-bacterial agents, steroids, painkillers etc 22 are all suitable. Optionally, the active ingredient 23 may be in delayed-release or controlled-release form. 24 25 The invention will now be further described with 26 reference to the following, non-limiting, examples and 27 Figures in which: 28 29 Fig. 1 H and E staining of 1240596-1 glass granule 30 intramuscular (six months). Magnification x 125. 31 Fig. 2 H and E staining of 1240596-2 glass granule 32 33 intramuscular (six months). Magnification x 125. 34 35 Fig. 3 H and E staining of 1240596-3 glass granule

11

1	intramuscular (six months). Magnification x 125.
2	-
3	Fig. 4 Neutrophil staining of muscle section
4	containing implant $1240596-1$ . Magnification x $125$
5	(black circles are air bubbles).
6	
7	Fig. 5 Macrophage staining of muscle section
8	containing implant $1240596-2$ . Magnification $x$
9	125.
10	
11	Example 1
12	
13	The CRGs will be implanted in vivo to assist in the
14	evaluation of attenuation of the solution rate of the
15	glass and to observe the acute tissue reaction at the
16	submucosal implant site.
17	
18	<u>Materials</u>
19	Two CRG compositions with slow solution rates (to be
20	decided) will be prepared as rough granules $200-300 \mu \mathrm{m}$
21	in diameter. The granules will be suspended in
22	glycerine BP, 8.5 ml glycerine to 10 g CRG. The
23	suspensions will be packaged 2.5 ml in syringes. The
24	syringes will be individually sealed in foil
25	polyester/polyester pouches and sterilised by $\gamma$
26	irradiation.
27	
28	Method
29	The anterior bladder wall of the anaesthetised model
30	(rabbit) is exposed and a small volume (0.5 ml) of CRG
31	implant is injected into the submucosa on the left and
32	right anterior bladder wall midway between the ureters
33	and the neck of the bladder. The implant should create
34	a small visible mound at the implant site. It is
35	suggested that the CRGs used at the left and right
	•

1 sites be of different solution rates, or that one of 2 the sites contains an existing "control" implant material for comparison (eg MACROPLASTIQUE (Trade Mark) 3 4 of Uroplasty, Inc ten animals would be required. 5 6 **Evaluation** 7 By placing the implants in the anterior bladder wall, 8 it should be possible to look at the implant on a 9 weekly basis using ultrasound. In addition, two animals would be sacrificed at two weeks, one month, 10 11 six months and twelve months. 12 13 The ultrasound examinations should look at the implant 14 material and any migration from the implant site should 15 be reported. Acute fibrous capsule formation should be 16 recorded. It may be possible to differentiate the CRG 17 and its dissolution over the more prolonged terms. 18 19 On sacrifice, tissue reactions and acute inflammation 20 should be recorded. Fibrous capsule development should 21 be noted and presence of CRG (and glycerol in early 22 stages) quantified for each implant site. Samples of 23 surrounding tissues should be removed for histological 24 examinations. 25 26 Results and Interpretations 27 An initial inflammatory response is anticipated at the 28 implant site. It is hoped that a collagen capsule will 29 form around the CRG granules. This capsule is expected 30 to reduce the solution rate of the glass. It will be 31 helpful to measure the attenuation of solution rate due 32 to reduced fluid transport within the capsule. By one 33 month surrounding tissue inflammations should have 34 subsided and histology should show normal cell 35 response. There should be no migration of the CRG

implant beads and the glycerol should be completely
removed within the first two weeks.

In each sacrifice group there should be at least one "control" implant. The tissue response of the control should be compared with the CRG implant results.

Example 2

9 <u>Materials and Methods</u>

10 Controlled release glasses (CRGs) were formulated as

11 follows:

		mole % concen	trations	
	Na₂O	CaO	P <sub>2</sub> O <sub>5</sub>	
1240596-1	5	48	47	
1240596-2	15	38	47	
1240596-3	25	28	47	

A granular diameter range of 53-1000  $\mu \mathrm{m}$  was used for all CRGs.

0.1 g samples of the CRGs listed above were sterilised by dry heat (190°C for 3 hours) before implantation into black and white hooded Lister rats (Liverpool strain). Two samples were implanted into each animal. Three animals were employed at a time period of six months. The implants were placed bilaterally into a pocket created in the dorsa-lumbar muscle region of the animal. At the six month time of explantation, the implant and surrounding tissue was removed from the sacrificed animal and frozen immediately. The frozen sample was sectioned at 7  $\mu$ m in a microtome cryostat. Analysis of the implant/tissue site was performed by staining the sample sections for various cytokines. A haematoxolin and eosin (H and E) stain was carried out

7	on each or cue	s bix recrieved samples, as well as			
2	neutrophil and	d macrophage staining.			
3	Immunohistoche	emical staining for ED1, ED2, CD4, CD8,			
4	Interleukin-1 $\beta$ (IL-1 $\beta$ ), IL-2, Major Histocompatibility				
5	_	class II, $\alpha$ - $\beta$ and Anti- $\beta$ antigens have			
6	been completed	d. These stains allow the tissue response			
7	_	t presence to be evaluated in the			
8	following man	-			
9	_				
10	H and E	Stains all viable cells and allows the			
11		tissue type and fibrous capsule to be			
12		easily identified by the characteristic			
13		structure shape of each tissue.			
14					
15	ED1	Recognises rat macrophages, monocytes			
16		and dendritic cells. Granulocytes are			
17		negative. The recognised antigen is			
18		predominantly located intracellularly,			
19		although some membrane expression			
20		occurs.			
21					
22	ED2	Recognises a membrane antigen on			
23		resident rat macrophages; monocytes,			
24		dendritic cells and granulocytes are			
25		negative. No other cell types but			
26		macrophages are positive for ED2, and it			
27		discriminates between thymic cortical			
28		(ED2+) and medullary macrophages (ED2-).			
29					
30	CD4	Expressed on most thymocytes and			
31		approximately two thirds of peripheral			
32		blood T cells. In humans and rats, CD4			
33		is expressed on monocytes and			
34		macrophages. CD4 is an accessory			
35		molecule in the recognition of foreign			

1		antigens in association with MHC class
2		II antigens by T cells.
3		
4	CD8	Expressed on most thymocytes and
5		approximately one third of peripheral
6		blood T cells, which constitute the CD4
7		negative cells. CD8 $\alpha$ is in all natural
8		killer (NK) cells in the rat.
9		
10	IL-1 $oldsymbol{eta}$	Expressed by B cells, macrophages and
11		monocytes and its mRNA is present in a
12		number of cells including T cells. In
13		addition to activating T and B
14		lymphocytes, interleukin-1 (IL-1)
15		induces several haematological and
16	•	metabolic changes typical of host
17		response to infection and injury. IL-1
18		is an endogenous pyrogen, producing
19		fever by its ability to increase
20		hypothalamic prostoglandin. IL-1 also
21		induces the release of several
22		lymphokines, interferons and colony
23		stimulating factors. With the exception
24		of skin keratinocytes, some epithelial
25		cells and certain cells in the central
26		nervous system, mRNA coding for IL-1 is
27		not observed in health in most other
28		cells.
29		
30	IL-2	More descriptively, T cell growth
31		factor, has promise as an immune
32		stimulant and an anti-tumour agent. IL-
33		2 recognises activated rat T cells but
34		not resting lymphocytes.
35		

16

1	MHC Class II	Expressed by dendritic cells, B cells,
2		monocytes, macrophages and some
3		epithelial cells. Expression is
4		increased by interferon $\alpha$ which also
5		induces expression on fibroblasts,
6		epithelial and endothelial cells.
7		
8	α-β	Detects an $\alpha$ - $\beta$ T cell receptor.
9		
10	Anti- $oldsymbol{eta}$	Directed at leucocytes. Also labels B
11		cells among thoracic duct lymphocytes
12		with little labelling in bone marrow and
13		none on thymocytes. Acts as an isotope
14		control.
15		
16		
17		
18		
19	Results and Di	scussion
20	The photograph	s in Figures 1-3 show H and E staining of
21	the I2405961-3	implants respectively. As can be seen
22	in these Figure	es, fibrous capsules have formed around
23	each glass gra	nule. Glass I240596-1 has the slowest
24	solution rate	as tested in-vitro, and this can be seen
25	-	so, as the sizes of the remaining glass
26	granules in the	e rat muscle after six months are
27		arger compared to the other two glass
28	•	hich both have faster solution rates
29	•	the fastest solution rate in-vitro).
30		g muscle tissue to the implant appears
31		res 4 and 5 show photographs of
32	~	ining of implant section I240596-1 and
33		ining of I240596-2 respectively. These
34		e typical of all the slides viewed, as
35	all six sectio	ns contained insignificant neutrophil and

17

macrophage presence in the tissue. In the photograph 1 2 of the neutrophil stained section, it can be seen that 3 there are several mast cells near the implant site and 4 throughout the tissue. This is expected in normal, 5 healthy muscle tissue. The lack of macrophages and 6 neutrophils indicates a lack of inflammatory response 7 to the implant, showing that after a six month period, 8 the glass granules appear to be accepted in-vivo. 9 10 The cytokine staining of the above antigens were all 11 negative, correlating with the absence of neutrophils 12 and macrophages in the tissue sections. Cytokines are 13 regulatory peptides that can be produced by virtually 14 every nucleated cell in the body, such as lymphocytes 15 and monocytes. Cytokines are generally not 16 constitutively produced, but are generated in 17 emergencies to contend with challenges to the integrity 18 of the host. Cytokines achieve these ends by 19 mobilizing and activating a wide variety of target cells to grow, differentiate and perform their 20 21 functions. This means that cytokines are key mediators 22 of immunity and inflammation. The insignificant 23 staining of the above indicates the acceptance of the 24 glass implant into the body and shows that the glass 25 presence is not inducing any inflammatory reaction in-26 vivo. 27 28 Conclusion 29 All the sections stained and viewed after the six month 30 period showed healthy, normal muscle tissue containing 31 a fibrous capsule coated glass granule. Staining of 32 various cytokines gave a negative result, indicating 33 the absence of inflammatory responses of the muscle 34 tissue with the glass presence after six months.

35

33

- 1 Example 3 2 Soft Tissue Response to Glycerol Suspended Controlled 3 Release Glass Particulates 4 This example investigated the soft tissue response of 5 glasses with a range of particulate sizes of different 6 dissolution rates, transported in a glycerol carrier. 7 8 Materials and Methods 9 The CRG was tested in particulate form of three 10 different compositions and two different particulate 11 sizes: X (200-300μm, 0.02 mg/cm<sup>2</sup>/hr solution rate), Y 12  $(200-300\mu\text{m}, 0.12 \text{ mg/cm}^2/\text{hr solution rate})$  and Z (<53 $\mu$ m, 13 0.34 mg/cm<sup>2</sup>/hr solution rate), all suspended in glycerol. A control sample of glycerol only was also 14 15 included in the experiment and was labelled sample W. 16 Samples weighing 0.1 grams of each of the CRG's in glycerol and glycerol only were sterilised by gamma 17 18 irradiation before implantation intramuscularly into 19 Wistar rats. Two samples were implanted into each 20 animal. Four animals at each time period of 2 days, 4 21 weeks, 9 weeks and 6 months were employed. 22 implants were placed bilaterally into a pocket created 23 in the dorso-lumbar muscle of the animal. At the time 24 of explantion, the implant and surrounding tissue was 25 removed from the sacrificed animal and snap frozen. A 26 microtome cryostat was used to cut 7µm thick serial 27 sections. Analysis of the implant/tissue site was 28 performed by specific staining the sample sections for 29 various cell types. Neutrophils and macrophages were 30 stained using enzyme histochemistry, ED1 (monocytes and 31 immature macrophages), ED2 (mature tissue macrophages), 32 CD4 (helper/inducer T-lymphocytes and macrophages), CD8
- 34 IL-2 (activated T-lymphocytes), Major
  35 Histocompatibility Complex (MHC) class II (activated

(suppressor/cytotoxic T-lymphocytes), interleukin- $1\beta$ ,

19

macrophages and activated B-lymphocytes),  $\alpha-\beta$  (T-

1

2 lymphocytes) and CD45RA (B lymphocytes) antibodies have 3 been used to immunohistochemically stain each sample. 4 5 Results and Discussion 6 Positive staining for neutrophils was observed after 2 day implantation with all of the materials. 7 8 neutrophils present were found in localised clusters near the implant site. However, neutrophils were not 9 10 seen in the tissue sections of each of the implanted 11 glasses or glycerol in the remaining time periods. 12 Mast cells were present in all tissue samples, but it 13 was noticed that an increased number of these cells 14 were present in clusters near the implanted glass in 15 sections containing glass X at 6 months, glass Y at 2 16 days and 6 months and glass Z at 4 weeks and 6 months. Enzyme staining and immunohistochemical staining both 17 confirmed the presence of macrophages in all sections 18 19 at all time periods except glass X at 6 months. 20 neutrophil presence at 2 days in all sections suggest 21 an acute inflammatory response. The absence of these 22 cells however in the remaining time periods indicate that this acute inflammation is quickly resolved. 23 24 However, the presence of macrophages in all samples at 25 all time periods except X at 6 months indicate an 26 ongoing chronic inflammatory response to the presence 27 of the implanted material. With glass X however, this 28 chronic inflammatory response appears to have been 29 resolved at 6 months. With one material, glass Z, 30 tissues necrosis in association with the glass at 4 31 weeks and 9 weeks has been observed. This study 32 demonstrates that particulate, degrading glass is 33 stimulating an inflammatory response in soft tissue of 34 time periods up to 6 months. It should be noted that 35 very small particulate fast degrading glass is leading

20

1 to tissue necrosis and should be further considered for 2 these applications. However, larger particulate, 3 slower degrading materials are demonstrating effective 4 potential for stress incontinence applications. 5 6 EXAMPLE 3 7 Inflammatory Response to Controlled Release Glass 8 Samples of a range of compositions of Controlled 9 Release Glasses (CRGs) in granular form were analysed 10 for the soft tissue response to determine their 11 biocompatibility. 12 13 Materials and Methods 14 The CRG was tested in granular form (53-1000µm) of 15 three different compositions: A (high in CaO, slow 16 solution rate), B (medium solution rate) and C (low in 17 CaO, fastest solution rate). Samples weighing 0.1 18 grams of each of the CRG's were sterilized by dry heat 19 (3hrs, 190°C) before implantation into black and white 20 hooded Lister rats. Two samples were implanted into 21 each animal. Three animals were employed at each time 22 period of 2 days, 1 week, 4 weeks, 8 weeks and 6 23 months. The implants were placed bilaterally into a 24 pocket created in the dorso-lumbar muscle of the 25 animal. At the time of explantion, the implant and 26 surrounding tissue was removed from the sacrificed 27 animal and snap frozen. The frozen sample was 28 sectioned at 7µm thickness in a microtome cryostat. 29 Analysis of the implant/tissue site was performed by 30 using different staining techniques. 31 Immunohistochemical staining using ED1 (monocytes and 32 immature macrophages), ED2 (mature tissue macrophages), 33 CD4 (helper/inducer T-lymphocytes and macrophages), CD8 34 (suppresser/cytotoxic T-lymphocytes), interleukin- $1\beta$ , 35 IL-2 (activated T-lymphocytes), Major

35

WO 98/44965 PCT/GB98/01017

21 1 Histocompatibility Complex (MHC) class II (activated 2 macrophages and activated T-lymphocytes),  $\alpha-\beta$  (T-3 lymphocytes) and CD45RA ( $\beta$ -lymphocytes) antibodies have 4 been performed. A haematoxylin and eosin (H and E) 5 stain was carried out on each of the retrieved samples. 6 Neutrophil and macrophage enzyme staining was also 7 performed. 8 9 Results and Discussion 10 The tissue response to the range of CRG's can clearly 11 be demonstrated as being different and dependant on the 12 materials, involving neutrophils, macrophages and mast 13 cells and not involving T or B lymphocytes. 14 15 Localised clusters of neutrophils were observed after 2 16 days implantation of each of the CRG's A, B and C. 17 However, neutrophils were not seen in the tissue 18 sections of each implanted glass in each of the 19 remaining time periods. 20 21 Mast cells were scattered throughout all tissue 22 sections as expected, but it was noticed that an 23 increased number of these cells were present in 24 clusters near the implant in sections containing CRG A 25 at 9 weeks and 6 months, and in CRG C at 2 days, 9 26 weeks and 6 months. 27 28 The most predominant cell type in all sections was the 29 macrophage confirmed by both enzyme staining and 30 immunohistochemistry. Macrophages were observed in all 31 of the sections for all of the time periods and were 32 positive for ED1, ED2 and MHCII antibodies. 33 presence of neutrophils at 2 days in all three glass 34 compositions indicate that an acute inflammatory

response has occurred. The absence of the neutrophils

22

at all subsequent time periods suggest that the acute inflammatory phase had resolved. However, the observation of macrophages throughout all time periods up to and including 6 months indicates continued stimulus by the materials of a chronic inflammatory phase response.

		23
1	CLA	IMS
2		
3	1.	A composition suitable for implantation in soft
4		tissue, said composition comprising particles of
5		biodegradable glass in a carrier medium.
6		
7	2.	A composition as claimed in Claim 1 wherein the
8		glass particles are irregularly shaped.
9		
10	3.	A composition as claimed in either one of Claims 1
11		and 2 wherein said particles have a diameter of
12		1000 μm or less.
13		
14	4.	A composition as claimed in any one of Claims 1 to
15		3 wherein said particles have a diameter of 300 µm
16		or less.
17		
18	5.	A composition as claimed in any one of Claims 1 to
19		4 wherein said particles have a diameter of 50 $\mu m$
20		to 100 µm.
21		
22	6.	A composition as claimed in any one of Claims 1 to
23		5 wherein said carrier medium is glycerol.
24		
25	7.	A composition as claimed in any one of Claims 1 to
26		6 wherein said carrier medium includes a
27		surfactant and/or a suspending agent.
28		
29	8.	A composition as claimed in any one of Claims 1 to
30		7 comprising glass particles formed from a
31		controlled release glass.
32		
33	9.	A composition as claimed in any one of Claims 1 to

9. A composition as claimed in any one of Claims 1 to
8 comprising glass particles formed from a watersoluble glass.

29 30

		24
1	10.	A composition as claimed in any one of Claims 1 to
2		9 comprising glass particles formed from a silver
3		containing glass.
4		
5	11.	Use of a composition as claimed in any one of
6		Claims 1 to 10 for augmentation of soft tissue.
7		
8	12.	Use as claimed in Claim 11 wherein said soft
9		tissue is the submucosa of the urethral sphincter.
10		
11	13.	A method of augmenting an area of soft tissue in a
12		body, said method comprising injecting a
13		composition as claimed in any one of Claims 1 to
14		10 into the soft tissue.
15		
16	14.	A method as claimed din Claim 13 which is used
17		augment soft tissue for cosmetic purposes.
18		
19	15.	A method as claimed in Claim 13 wherein said soft
20		tissue is the submucosa of a wall of a body organ.
21		
22	16.	A method of combatting vesicoureteric reflux by
23		injecting a composition as claimed in any one of
24		Claims 1 to 10 into the bladder submucosa close to
25		the urethral orifice such that urine is unable to
26		pass up the ureter upon contraction of the
27		bladder.
28		

1/3

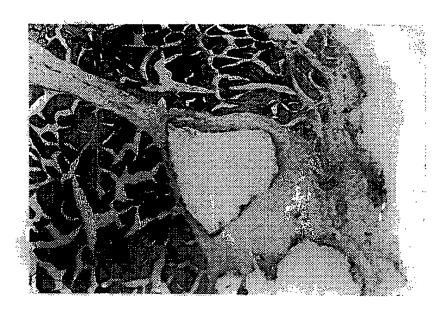


Fig. 1



Fig. 2 **SUBSTITUTE SHEET (RULE 26)** 

# BEST AVAILABLE COPY PCT/GB98/01017

2/3

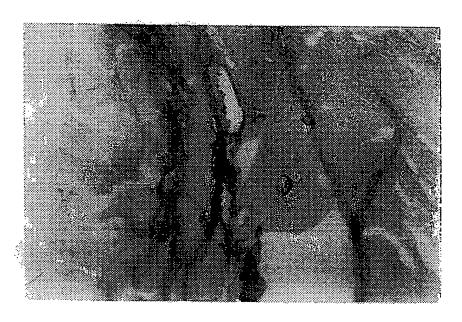


Fig. 3

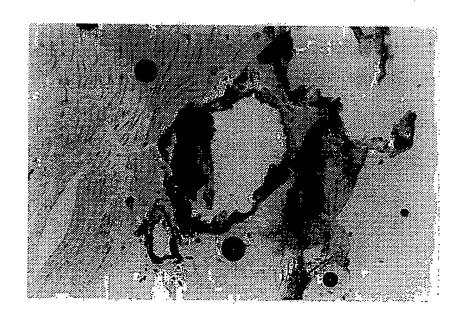


Fig. 4
SUBSTITUTE SHEET (RULE 26)

# BEST AVAILABLE COPY

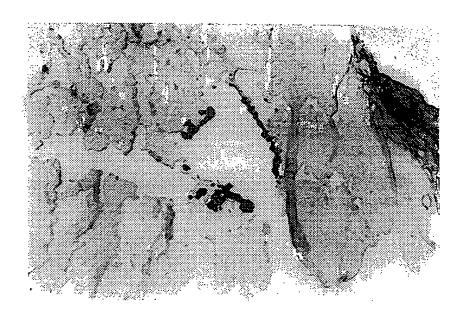


Fig. 5

In: Itional Application No

			1 017 40 307	01017
A. CLASSI IPC 6	FICATION OF SUBJECT MATTER A61L27/00			
According to	o International Patent Classification (IPC) or to both national classifica	ition and IPC		
	SEARCHED			
	ocumentation searched (classification system followed by classification A61L C03C	n symbols)		
Documentat	tion searched other than minimum documentation to the extent that su	uch documents are inclu	ided in the fields sea	erched
Electronic d	ata base consulted during the international search (name of data bas	se and, where practical,	search terms used)	
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT			
Category °	Citation of document, with indication, where appropriate, of the rele	vant passages		Relevant to claim No.
Х	US 5 204 382 A (WALLACE DONALD G 20 April 1993 see column 7, line 36 - line 47;	ŕ		1,3-6, 11-16
Х	WO 93 16658 A (JSF CONSULTANTS LTD) 2 September 1993 see page 6, line 17 - line 28; claims		1,6,11, 12	
Υ	WO 93 15721 A (HUBBARD WILLIAM G) 19 August 1993 see claims		1-16	
P,Y	WO 97 33632 A (GILTECH LTD ;GILCHRIST EILIDH (GB); GILCHRIST THOMAS (GB)) 18 September 1997 see claims; examples		1-16	
	_ <del></del>	-/		
		,		
	her documents are listed in the continuation of box C.	X Patent family	members are listed i	n annex.
° Special ca	tegories of cited documents:	"T" later document put		
"A" document defining the general state of the art which is not considered to be of particular relevance invention  "E" earlier document but published on or after the international			ed the principle or the	eory underlying the
filing date  "X" document of particular relevance; the cannot be considered novel or cannot be c			ered novel or cannot	be considered to
which is cited to establish the publication date of another citation or other special reason (as specified)  "Y" document of particular relevance; the cannot be considered to involve an i		ular relevance; the c	laimed invention	
"O" document referring to an oral disclosure, use, exhibition or other means ments, such combined with one or ments, such combination being obvious "P" document published prior to the international filling date but		ined with one or mo	re other such docu-	
later than the priority date claimed "&" document member of the same patent			amily	
	August 1009		ne international sea	rch report
	August 1998	11/08/1		
Name and m	nailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk	Authorized officer		
	Tel. (+31-70) 340-2040, Tx. 31 651 epo ni, Fax: (+31-70) 340-3016	ESPINOS	A, M	

1

In utional Application No PCT/GB 98/01017

	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	 
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 91 17777 A (UNIV FLORIDA) 28 November 1991 see claims	1-16
A	WO 90 08470 A (GILTECH LTD) 9 August 1990	
A	WO 96 24364 A (GILTECH LTD ;HEALY DAVID MICHAEL (GB); GILCHRIST THOMAS (GB)) 15 August 1996	
	·	

.ernational application No.

PCT/GB 98/01017

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X Claims Nos.: 13-16 because they relate to subject matter not required to be searched by this Authority, namely:  Remark: Although claims 13-16  are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest  The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.

Information on patent family members

In ational Application No PCT/GB 98/01017

Patent doo cited in sear		Publication date	Patent family member(s)		Publication date
US 5204:	382 A	20-04-1993	AU	3619693 A	13-09-1993
			CA	2128783 A	02-09-1993
			EP	0627899 A	14-12-1994
			JP	7504106 T	11-05-1995
			WO	9316657 A	02-09-1993
			ÜS	5352715 A	04-10-1994
WO 9316	558 A	02-09-1993	US	5480644 A	02-01-1996
			AU	674308 B	19-12-1996
			AU	3785393 A	13-09-1993
			CA	2131021 A	02-09-1993
			EΡ	0627900 A	14-12-1994
	•		JP	2711181 B	10-02-1998
			JP	7505146 T	08-06-1995
			US	5490984 A	13-02-1996
WO 9315	721 A	19-08-1993	AU	3612593 A	03-09-1993
			CA	2129993 A	19-08-1993
			EP	0631499 A	04-01-1995
			JP	6506862 T	04-08-1994
			NO	942966 A	10-08-1994
•			NZ	249381 A	21-12-1995
			SG	47024 A	20-03-1998
		·	ZA	9300506 A	11-05-1994
WO 97336	532 A	18-09-1997	AU	2032997 A	01-10-1997
WO 9117	777 A	28-11-1991	NON		
WO 90084	170 A	09-08-1990	AT	150935 T	15-04-1997
			DE	69030374 D	07-05-1997
			DE	69030374 T	16-10-1997
			DK	455706 T	13-10-1997
			EP	0455706 A	13-11-1991
			ES	2099708 T	01-06-1997
			JP	4503018 T	04-06-1992
			US	5470585 A	28-11-1995
WO 96243	364 A	15-08-1996	AU	4724196 A	27-08-1996
			CA	2212168 A	15-08-1996

Information on patent family members

in tilonal Application No PCT/GB 98/01017

Patent document cited in search report			Publication date	Patent family member(s)		Publication date
WO	9624364	Α	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	EP	0809506 A	03-12-1997
					·	
					•	
				•	-	
			,			
			•			
						•